

DISTRIBUTION OF DF³²P IN MOUSE ORGANS—I. THE EFFECT OF ROUTE OF ADMINISTRATION ON INCORPORATION AND TOXICITY

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Abstract—The LD₅₀ of di-isopropyl phosphorofluoridate (DFP) for mice by the intraperitoneal route is found to be 6.8 mg/kg while that by the subcutaneous or intravenous route is found to be much lower (3.8 and 3.4 mg/kg respectively). When radioactive di-isopropyl phosphorofluoridate (DF³²P) is injected to mice or rats by the intraperitoneal route there is a higher uptake of radioactivity in the liver with a corresponding reduced incorporation in other organs than when the administration is by the subcutaneous or the intravenous routes. Death occurs when the concentration of DF³²P reaches a certain critical value in some organs including the brain whatever the route employed. A large part of the detoxication of DFP which is known to take place in the liver is thus due to its absorption by the liver esterases whose inhibition obviously does not contribute materially to the overall toxicity.

THOUGH it is well known that organophosphorus esters owe their toxicity to their ability to combine irreversibly with the active site of acetylcholinesterase it is difficult to establish any predictable pattern correlating the chemical structure of an organophosphate with its lethal effect.¹ Even within a given species the LD₅₀ is dependent upon the route of administration. These differences are sought to be explained by many factors the most important being the extent of detoxication taking place in the animal system.

In a previous paper² it was shown that the enzyme DFPase which hydrolyzes di-isopropyl phosphorofluoridate (DFP) is present only in small amounts in rat and rabbit tissues. It was also reported^{3, 4} that liver microsomes contain large amounts of non-specific esterases which incorporate DFP. In the present work data are presented to show that the detoxication occurring in the liver is mainly due to the absorption of DFP by microsomal esterases whose inhibition obviously does not contribute to the overall lethal effect. The LD₅₀ of DFP by the intraperitoneal (i.p.) route is found to be much higher than those by the subcutaneous (s.c.) and the intravenous (i.v.) routes. This excess dose is correlated to the higher uptake of DFP in the liver when the injection is given by the i.p. route. There is a correspondingly reduced uptake in the other organs which are probably the foci of lethal action since death occurs when the DFP concentration reaches a certain critical value in these organs whatever is the route of administration. Radioactive di-isopropyl phosphorofluoridate (DF³²P) admixed with DFP is used in these studies.

MATERIALS AND METHODS

The samples of DFP used in this work were obtained from the Research Institute of National Defence, Department 1, Sundbyberg 4, Sweden. Stock solutions were

prepared which contained 25 mg/g in dry propylene glycol. DF^{32}P with a specific activity of 300 mc/g was obtained from the Radiochemical Centre, Amersham. Stock solutions of this contained 1.85 mg ($10\ \mu\text{moles}$)/g in propylene glycol. The solutions were preserved at -16° and quantities were accurately weighed out and dissolved in 0.9% sodium chloride solution just before injections. When mixtures of DFP and DF^{32}P were to be used 20 per cent of the organophosphate was derived from the DF^{32}P stock solution. It was found that DF^{32}P usually contained 8–15 per cent of a radioactive impurity which was not extractable by ether.

Male albino mice weighing 22–25 g were used throughout. Animals of uniform weight (± 1 g) were chosen for any series of experiments and a constant volume of solution (0.08–0.1 ml) was injected to each mouse. The s.c. injections were given under the loose skin in the back and the i.v. injections were in the tail vein. Since the rate of i.v. injection is known to influence toxicity⁵ these were administered at a controlled rate of 0.1 ml/20 sec. The LD_{50} values were calculated by Thompson's method⁶ using the tables compiled by Weil.⁷ Four groups of 6 animals each were usually employed for each assay, the dosages being so arranged in a geometric progression that their logarithms differed by 0.025 or 0.03. A 24-hr observation period was allowed for death or survival.

For the determination of radioactive DFP incorporated in various organs, groups of 5 or more animals were injected with the necessary amount of DFP + DF^{32}P mixture at suitably arranged time intervals and each animal was killed individually exactly 30 min after the injection. Sacrifice was by stunning followed by exsanguination to remove as much of the blood as possible. The liver, kidneys and lungs were quickly removed, cut into small bits and rinsed with physiological saline and finally homogenized in a Potter–Elvehjem type of homogenizer. The volume was made up to 25 ml with saline and 1 ml aliquots of the uniform suspensions were pipetted into glass cups and dried after the addition of a drop of 4 N NaOH. The brain tissue was prepared by dissolving in 6 ml of hot 1 N NaOH. Each animal was worked up separately and radioactivity measurements were taken in triplicate in a Robot Equipment (LKB-Produkter, Stockholm) fitted with a Tracerlab Compumatic Scaler. The lung tissue was often contaminated with blood especially at dose levels approaching the LD_{50} .

Studies on rats were performed with male animals weighing about 300 g. The injection volume in these cases was 1 ml per rat.

RESULTS

LD_{50} of DFP by various routes

This was determined using DFP unadmixed with DF^{32}P . Two different batches of the substance were used. The results are given in Table 1. Most of the deaths occurred in the first few hours, late deaths being more frequent in i.v. studies.

Incorporation of DF^{32}P administered s.c. and i.p.

Quantities of DFP and DF^{32}P were weighed out and dissolved in the necessary volume of saline so that 0.1 ml contained the required amount of the organophosphate to give dosages of 0.5, 1.0, 2.0 and 4.0 mg/kg body weight. In the last dose level there were a few deaths by the s.c. route but only the survivors at 30 min were analysed. Since the LD_{50} by the i.p. route has been found to be 6.8 mg/kg groups of mice were

TABLE 1. LD₅₀ OF DFP FOR MICE BY DIFFERENT ROUTES OF ADMINISTRATION

| Route | LD ₅₀ (mg/kg) | Range (95% confidence limits) (mg/kg) |
|-------|-----------------------------|---|
| i.p. | 6.7 | 6.0-7.4 |
| | 6.8 | 6.4-7.1 |
| s.c. | 3.6 | 3.5-3.7 |
| | 3.9* | 3.7-4.0 |
| i.v. | 3.2 | 3.0-3.4 |
| | 3.5 | 3.3-3.8 |

LD₅₀ was determined using 4 groups of 6 mice each (5 mice in the case of series marked *). DFP solution was freshly prepared by weighing accurate amounts of the stock solution in propylene glycol and dissolving in the necessary volume of 0.9 per cent saline. The dosages were in geometric progression, their logarithms differing by 0.025 or 0.03. The LD₅₀s were calculated according to the method of Thompson⁶ using the tables compiled by Weil.⁷ A 24-hr observation period was allowed for death or survival.

also injected with the DFP + DF³²P mixture at dose levels of 6.0, 6.5 and 7.0 mg/kg by this route. There were a few deaths at the last dose level. The results are given in Table 2. The numbers in brackets in the following text refer to the order numbers of experiments in the table.

It is seen that at any dose level the uptake of DF³²P in the liver is much higher by the i.p. than by the s.c. route (1-8). All the differences are statistically significant. It is also observed that at 4.0 mg/kg (s.c.) which is the approximate LD₅₀ by the s.c. route there is less DF³²P in the liver than when the same amount is injected i.p. (4). This indicates that the DF³²P content of the liver does not determine death. On the other hand, there is actually an un-utilized detoxicating potential in the liver at the time of death of the animal by 4.0 mg/kg s.c.

The uptake of DF³²P in the kidney follows a reverse order, the amounts incorporated being less by the i.p. route than by the s.c. route (9-12) at sublethal doses. However, when the dose level approaches the LD₅₀ the differences are hardly significant (13-16). In the case of the lung the incorporation is, in general, less in the case of i.p. injections (17-21). However, in the course of the experiments it was observed that the lung tissue was often contaminated with large amounts of blood which was difficult to be rinsed out by cutting the tissue in small bits in saline. Erratic values for radioactivity were obtained in such cases and these were more frequent as the DFP dose level approached the LD₅₀.

The incorporation of DF³²P in the brain follows the same pattern as in the kidneys but is more consistent. The differences between the s.c. and i.p. values are highly significant up to 4.0 mg/kg which is the approximate LD₅₀ (s.c.) after making allowance for the slight impurity in the DF³²P (25-28). At this level by the i.p. route it is seen that the incorporation in the brain is considerably less. When the i.p. dose is progressively increased from 6.0 mg/kg the amount of DF³²P in the brain approaches a value which is very near that of the 4.0 mg (s.c.) value (29-32). This indicates that at the time of death the DFP content of the brain is the same whatever the route of administration.

TABLE 2. INCORPORATION OF DF³²P IN MOUSE ORGANS WHEN ADMINISTERED BY THE s.c. AND i.p. ROUTES

| No. | Organ | Dose DFP + DF ³² P mg/kg | Activity injected | Activity incorporated in the organs, s.e.m. and number of animals | | s.c./i.p. ratio | P |
|-----|--------|--|----------------------|--|-----------------|--------------------|--------|
| | | | | s.c. route | i.p. route | | |
| 1 | Liver | 0.5 | 190 | 32 ± 0.9 (5) | 69 ± 1.1 (5) | 0.46 | <0.001 |
| 2 | | 1.0 | 380 | 101 ± 2.6 (5) | 147 ± 8.2 (4) | 0.69 | <0.001 |
| 3 | | 2.0 | 760 | 186 ± 6.4 (5) | 217 ± 10.2 (5) | 0.86 | <0.001 |
| 4 | | 4.0 | 1520 | 259 ± 15.7 (5) | 319 ± 10.2 (5) | 0.81 | <0.001 |
| 5 | | 4.0 | 294 | 60 ± 3.6 (5) | | | |
| 6 | | 6.0 | 441 | | 75 ± 6.4 (3) | 0.80 | <0.01 |
| 7 | | 6.5 | 520 | | 76 ± 3.5 (5) | 0.79 | <0.001 |
| 8 | | 7.0 | 553 | | 83 ± 3.4 (4) | 0.72 | <0.001 |
| 9 | Kidney | 0.5 | 190 | 15 ± 1.2 (5) | 8 ± 1.2 (5) | 1.88 | <0.001 |
| 10 | | 1.0 | 380 | 24 ± 1.9 (5) | 18 ± 1.5 (5) | 1.33 | <0.005 |
| 11 | | 2.0 | 760 | 35 ± 1.7 (5) | 27 ± 3.2 (3) | 1.30 | <0.005 |
| 12 | | 4.0 | 1520 | 78 ± 7.8 (5) | 66 ± 2.4 (5) | 1.18 | <0.02 |
| 13 | | 4.0 | 294 | 17 ± 1.6 (7) | 10 ± 0.7 (5) | 1.70 | <0.001 |
| 14 | | 6.0 | 441 | | 20 ± 3.5 (5) | 0.85 | >0.1 |
| 15 | | 6.5 | 520 | | 16 ± 1.8 (4) | 1.06 | >0.4 |
| 16 | | 7.0 | 553 | | 18 ± 1.9 (3) | 0.94 | >0.4 |
| 17 | Lung | 0.5 | 190 | 4.4 ± 0.2 (4) | 4.3 ± 0.3 (4) | 1.02 | >0.5 |
| 18 | | 1.0 | 380 | 5.5 ± 0.8 (5) | 5.8 ± 0.5 (5) | 0.95 | >0.5 |
| 19 | | 2.0 | 760 | 7.7 ± 0.7 (5) | 5.9 ± 0.3 (5) | 1.31 | <0.005 |
| 20 | | 4.0 | 1520 | 9.3 ± 1.0 (3) | 7.9 ± 0.2 (5) | 1.18 | <0.01 |
| 21 | | 4.0 | 294 | 2.6 ± 0.3 (7) | 1.8 ± 0.2 (5) | 1.44 | <0.005 |
| 22 | | 6.0 | 441 | | 1.8 ± 0.3 (6) | 1.44 | <0.005 |
| 23 | | 6.5 | 520 | | 2.3 ± 0.3 (5) | 1.13 | >0.2 |
| 24 | | 7.0 | 553 | | 2.4 ± 0.5 (4) | 1.08 | >0.4 |
| 25 | Brain | 0.5 | 190 | 0.62 ± 0.03 (5) | 0.30 ± 0.02 (5) | 2.07 | <0.001 |
| 26 | | 1.0 | 380 | 1.04 ± 0.04 (5) | 0.54 ± 0.05 (5) | 1.93 | <0.001 |
| 27 | | 2.0 | 760 | 1.92 ± 0.09 (5) | 0.93 ± 0.12 (5) | 2.06 | <0.001 |
| 28 | | 4.0 | 1520 | 3.82 ± 0.06 (5) | 1.89 ± 0.10 (5) | 2.02 | <0.001 |
| 29 | | 4.0 | 294 | 1.10 ± 0.07 (7) | 0.69 ± 0.07 (4) | 1.59 | <0.001 |
| 30 | | 6.0 | 441 | | 0.71 ± 0.16 (6) | 1.55 | <0.001 |
| 31 | | 6.5 | 520 | | 1.04 ± 0.09 (4) | 1.06 | >0.3 |
| 32 | | 7.0 | 553 | | 1.28 ± 0.24 (4) | 0.86 | >0.05 |

Values are expressed in count/min $\times 10^{-3}$ in whole organs. Albino mice of 22–25 g were used in these experiments but in any one series animals of uniform weight (± 1 g) were chosen and a constant volume of DFP + DF³²P solution (0.08 to 0.1 ml) was injected. This solution was prepared fresh by weighing out quantities of a stock solution of DFP (25 mg/g) and DF³²P (1.85 mg/g) in propylene glycol and dissolving in the required volume of saline to give the necessary dosage. 20 per cent of the total DFP was derived from the DF³²P stock solution. Five or more animals were used for each determination but each animal was worked up separately on a previously worked out time schedule for injection and sacrifice. To minimise errors due to spontaneous hydrolysis of DFP in aqueous medium fresh quantities of the stock solutions were weighed and diluted for every 20 animals (about 100 min). The interval between injection and sacrifice was 30 min in all cases. Other details are given under Materials and Methods.

Incorporation of DF³²P in organs when administered s.c. and i.v.

The amounts of DF³²P incorporated in the organs when graded doses of DFP + DF³²P are injected by the s.c. and i.v. routes are given in Table 3. As in the case of the s.c. and i.p. experiments three i.v. dose levels near the LD₅₀ were also included in the series, viz., 3.2, 3.4 and 3.6 mg/kg, but all the mice administered 3.6 mg/kg i.v. died within 30 min and these values are therefore not presented.

In general, the differences between the s.c. and i.v. values are less significant than those in the s.c. and i.p. series. It is also noteworthy that the LD₅₀s by these routes lie close together viz., 3.8 and 3.4 mg/kg respectively (Table 1).

TABLE 3. INCORPORATION OF DF³²P IN MOUSE ORGANS BY THE s.c. AND i.v. ROUTES

| No. | Organ | Dose DFP + DF ³² P (mg/kg) | Activity injected | Activity incorporated in the organs, s.e.m. and number of animals | | s.c./i.v. ratio | P |
|-----|--------|--|----------------------|--|------------------|--------------------|--------|
| | | | | s.c. route | i.v. route | | |
| 33 | Liver | 0.5 | 37 | 6.8 ± 0.71 (6) | 9.5 ± 1.12 (6) | 0.72 | <0.005 |
| 34 | | 1.0 | 74 | 18.6 ± 1.33 (6) | 22.9 ± 1.15 (6) | 0.81 | <0.001 |
| 35 | | 2.0 | 148 | 27.6 ± 3.40 (6) | 36.1 ± 2.39 (6) | 0.76 | <0.005 |
| 36 | | 4.0 | 296 | 46.9 ± 3.49 (5) | | | |
| 37 | | 3.2 | 237 | | 50.9 ± 2.43 (5) | 0.92 | >0.05 |
| 38 | Kidney | 3.4 | 252 | | 48.9 ± 2.33 (7) | 0.96 | >0.1 |
| 39 | | 0.5 | 37 | 2.4 ± 0.15 (6) | 2.7 ± 0.42 (6) | 0.89 | >0.2 |
| 40 | | 1.0 | 74 | 3.7 ± 0.27 (6) | 2.8 ± 0.13 (6) | 1.32 | <0.001 |
| 41 | | 2.0 | 148 | 5.2 ± 0.80 (6) | 4.5 ± 0.27 (6) | 1.16 | >0.1 |
| 42 | | 4.0 | 296 | 14.8 ± 3.29 (5) | | | |
| 43 | Lung | 3.2 | 237 | | 16.0 ± 2.71 (5) | 0.93 | >0.7 |
| 44 | | 3.4 | 252 | | 11.1 ± 0.96 (7) | 1.33 | <0.05 |
| 45 | | 0.5 | 37 | 0.75 ± 0.06 (6) | 0.83 ± 0.10 (6) | 0.90 | >0.1 |
| 46 | | 1.0 | 74 | 0.89 ± 0.06 (6) | 0.75 ± 0.05 (6) | 1.19 | <0.005 |
| 47 | | 2.0 | 148 | 0.92 ± 0.15 (6) | 1.19 ± 0.12 (6) | 0.77 | <0.02 |
| 48 | Brain | 4.0 | 296 | 2.72 ± 0.28 (5) | | | |
| 49 | | 3.2 | 237 | | 1.47 ± 0.15 (5) | 1.85 | <0.001 |
| 50 | | 3.4 | 252 | | 1.67 ± 0.12 (7) | 1.63 | <0.001 |
| 51 | | 0.5 | 37 | 0.13 ± 0.006 (6) | 0.14 ± 0.005 (6) | 0.93 | <0.05 |
| 52 | | 1.0 | 74 | 0.25 ± 0.018 (6) | 0.28 ± 0.006 (6) | 0.89 | <0.02 |
| 53 | | 2.0 | 148 | 0.33 ± 0.030 (6) | 0.41 ± 0.030 (6) | 0.80 | <0.005 |
| 54 | | 4.0 | 296 | 0.80 ± 0.160 (5) | | | |
| 55 | | 3.2 | 237 | | 0.80 ± 0.070 (5) | 1.00 | >0.9 |
| 56 | | 3.4 | 252 | | 0.79 ± 0.040 (7) | 1.01 | >0.8 |

Values are expressed in count/min $\times 10^{-3}$. The details are the same as given under Table 2 and Materials and Methods. The i.v. injections were given in the tail vein keeping an approximate rate of 0.1 ml/20 sec.

There are significant differences in the DF³²P uptake in the liver at sublethal doses (33–35) the incorporation being higher by the i.v. route. As the dosages approach the LD₅₀ (36–38) the differences are no longer significant. The values for the kidneys and lungs do not show consistency, the s.c./i.v. ratio being found to lie on either side of unity. As pointed out earlier the lung tissue gives erratic values at near LD₅₀ levels. As in the case of the s.c.–i.p. comparison experiments the DF³²P uptake in the brain shows a remarkable consistency. There are statistically significant differences between the s.c. and i.v. values at sublethal dose levels (51–53), the i.v. values being higher. At doses near the LD₅₀ the DF³²P content of the brain reaches an almost identical value by either mode of administration (54–56).

Dose-uptake curves

If the uptake of DF³²P (Tables 2 and 3) is plotted against the logarithm of the doses from 0.5 to 4.0 mg/kg, it is possible to obtain a straight line curve for the liver up to 4.0 mg/kg. However, in the case of the other organs the curves are linear only up to 2.0 mg/kg after which there is a sudden steep rise in the slope. For economy of space these curves are not given but attention is drawn to the high values of item numbers 12, 28, 42, 48 and 54 in Tables 2 and 3. A possible explanation can be that the detoxication capacity of the liver is getting rapidly exhausted in the region 2.0 to 4.0 mg/kg and this results in an abrupt overflow into the other organs.

Excretion of DF³²P after administration by the s.c. and i.p. routes

An organophosphate has to pass through the liver first if it is injected by the i.p. route. It can therefore be expected that its enzymic degradation will be much higher if administered by this route. Adie⁸ found that there is a relationship between the sarinase level of the rabbit liver and the dose of sarin needed to kill the animal. To determine if the higher LD₅₀ of DFP by the i.p. route is due to its higher enzymic degradation by the liver DFPase, excretion studies were conducted on 2 sets of rats injected DF³²P by the s.c. and i.p. routes respectively. A higher excretion of radioactive material should

TABLE 4. INCORPORATION OF EXCRETION OF DF³²P IN RATS

| Organ | Interval between injection and sacrifice (hr) | Radioactivity* when injected | | |
|--------------------------|---|------------------------------|--------------|-----------|
| | | (s.c. route) | (i.p. route) | s.c./i.p. |
| Liver | 1 | 277 | 371 | 0.75 |
| | 24 | 188 | 303 | 0.62 |
| Kidney | 1 | 162 | 81 | 2.00 |
| | 24 | 58 | 41 | 1.41 |
| Lung | 1 | 27 | 28 | 0.96 |
| | 24 | 17 | 18 | 0.94 |
| Brain | 1 | 4.1 | 2.3 | 1.78 |
| | 24 | 2.3 | 1.1 | 2.09 |
| Excreta (urine + faeces) | 24 | 2190 | 2212 | 0.99 |

* The values are expressed in count/min $\times 10^{-3}$. Each figure represents the average of 2 rats of weight 290–310 g. The rats were injected 1 ml each of a DF³²P solution (0.8 mg of DFP/kg) which contained 3.53×10^6 count/min of radioactivity. The animals used in the excretion studies were allowed free access to food and water. The 24-hr values for the organs pertain to these animals.

result in the set of rats injected i.p. as the DF³²P, once it is hydrolyzed to di-isopropyl phosphate by the DFPase, will not bind to tissues⁹ and will be excreted. As results in Table 4 show this does not seem to take place. The excretion of radioactive phosphorus is almost identical in both the sets and it can therefore be concluded that the higher uptake of DF³²P in the liver by the i.p. mode of administration is compensated by a lower uptake in the other organs and that there is no significant overall increase in the degradation of DF³²P if it is injected intraperitoneally.

DISCUSSION

From the foregoing results it is apparent that the wide gap between the LD₅₀s of DFP by the s.c. and i.p. routes of administration can be explained by the differences in the distribution ratio of the organophosphate in the organs when administered by these routes. It is found that more DF³²P is incorporated in the liver when the injection is by the i.p. route with a correspondingly reduced uptake in the other organs which are probably the vital targets of lethal action (Table 2). The higher uptake of DF³²P in the liver by the i.p. route is predictable since the liver has a high content of DFP-susceptible esterases^{3, 4} and the injected substance has to pass through this organ. As stable phosphorylated enzymes are formed³ the initial quantities of the injected DF³²P will be used up by the microsomal esterases, only the overflow reaching the other organs. It is clear that the liver itself is not a vital organ involved in the lethal effects of DFP as it is possible to have a high concentration of DFP in the liver without

causing death (see item 4 in Table 2). On the other hand, the liver seems to act as a buffer absorbing the organophosphate in the esterases, whose inhibition, whatever may be its long-term effect, does not apparently contribute to the acute toxicity. The detoxication effected by the liver is thus mainly by the absorption of DFP by the microsomal esterases and enzymic degradation seems to play only a minor role since there is no increased excretion of radioactivity when DF³²P is injected i.p. (Table 4). This confirms our earlier finding that the enzyme DFPase has only a low activity in the animal system.² An abrupt rise in the uptake of DF³²P by the kidney, lung and the brain in the region of the LD₅₀ also points to a buffering effect by the liver at the initial stages.

It is likely that the liver acts in a similar way in modifying the effective doses of many other active substances. Especially of interest will be a re-assessment of the action of oxime antidotes in the light of the above findings as these are most often administered by the i.p. route in experimental work. Studies along these lines are in progress.

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